CONNECTIVE TISSUE AND THE ATTACHMENT OF PIN BONE IN SALMON AND COD

Why are the pin bones so firmly attached?

Mona E. Pedersen
What is the problem?

– The pin bones are difficult to remove early post-mortem from the filet.

– When removed either the filet is damaged, or the pin bones break inside the muscle.

– The pulling force of pin bones decreases post mortem, differ between anterior and posterior position in the fish, and is higher in cod compared to salmon (Leif Akse et al, Fiskeriforskning, Rapport 15/2002).

– Little is known about how pinbones are attached to the muscle.
Connective tissue = a complex structural network

Extracellular matrix (proteoglycans, collagens and glycoproteins)

Adhesion proteins (syndecans, glypicans, integrins)

Cells (fibroblasts, fat cells, immune cells)

Enzymes (MMPs, serine proteases, aggrecanases, cathepsines etc.)
Strong interaction of carbohydrates and proteins
Aim of the study

- Characterize the structure of the attachment
- Identify connective tissue components in the structure
- Study enzymes and the degradation process *post mortem*
Sampling

- Salmon and cod
- 0h, 6h, 12h, 24h, 48h, 3 days and 5 days storage
- Dissected 6000 pinbones from anterior and posterior position in the filet
- Either fixed or frozen in liquid nitrogen before further analysis
Methods used in the study

- **Microarray**: Screening of components in the structure. What is expressed of connective tissue components, adhesion proteins, enzymes?

- **Histology**: Study structure, localization of relevant proteins and degradation of the structure *post mortem*

- **Zymography**: Identify enzymes and their activity *post mortem*

- **Proteomics**: Identify relevant proteins that are changed (0h and 5d). Screening of proteins

- **Western blotting**: Verify changes of relevant single proteins during storage period
Interphase connective tissue - bone
Gene expression analysis

- Pooled samples of the two most anterior and posterior pin bones from four fish were selected for microarray gene expression analysis.

- Pooled samples of muscle from all four fish were used as reference in the analysis.

- Comparison of gene expression profile:
  - Pin bone vs. muscle
  - Anterior pin bone vs. posterior pin bone
Results – cod

- > 2000 differentially expressed genes between pin bone and muscle
- Enrichment analysis of differentially expressed genes

Examples of genes:
- Extracellular matrix: collagen I, IV collagen V, collagen XI, collagen XII, decorin, laminin
- Lipid metabolism: fabp, lipase, acyl CoA synthetase, acyl CoA dehydrogenase
- Protease: MMP13, calpain, cathepsin F, cathepsin H, serine-protease, elastase
- Adhesion: Integrins
Results – salmon

- >193 differentially expressed genes in pin bone vs. muscle
- Examples of genes:
  - Extracellular matrix: collagen I, collagen III, collagen X, collagen XV, lumican, transgelin,
  - Proteases: collagenase, cathepsin K, MMP2, TIMP2, serine protease
Results – cod and salmon

- Generally higher gene expression levels in anterior vs posterior pin bones of both species
- Different extracellular matrix composition between the two species
The connective tissue is rich of elastin, proteoglycans and collagens

- Dark colour: Elastin
- Brown: Muscle

- Dark colour: Proteoglycans
- Light blue: Muscle
Tight connection between bone, connective tissue and muscle!!
But what happens during storage?
Elastin fibres are broken
Threadlike structures containing proteoglycans
Localization of collagen X and I in the splitting area
The network of carbohydrate binding protein and collagen in the splitting area
Metalloproteases are active right after slaughter in salmon.
Metalloproteases activity is high during the storage period in cod.
Different types of degrading enzymes are active
Conclusions

- The connective tissue is important!
- Differences between muscle and pin bone area
- Differences between salmon and cod
- Differences in enzymatic profile between salmon and cod
- The connective tissue is broken during storage into threadlike structures
- It is the attachment between bone and connective tissue that is degraded *post mortem*
Further work

• Histology of salmon and relevant candidate proteins identified by microarray analysis

• Study changes in proteins by proteomics during storage (0h and 5d)

• Study changes of relevant single proteins by western blotting during the storage period

• Identification of MMP types and localization
How can we use this information?

• When we know the pin bone biology we can:
  – Develop better methods for pin bone removal
  – Predict and examine which external factors that can be important
  – Optimize the pulling force
Acknowledgments

Sissel B. Rønning
Kristin Hollung
Tone-Kari Østbye
Thomas Larsson
Grethe Enersen
Vibeke Høst
Karen Sanden
Karin Solgård

This project is funded by FHF